



Soil Science and Plant Nutrition

ISSN: 0038-0768 (Print) 1747-0765 (Online) Journal homepage: https://www.tandfonline.com/loi/tssp20

Decomposition and CO₂-C evolution of okara, sewage sludge, cow and poultry manure composts in soils

Michael Miyittah & Kazuyuki Inubushi

To cite this article: Michael Miyittah & Kazuyuki Inubushi (2003) Decomposition and CO₂-C evolution of okara, sewage sludge, cow and poultry manure composts in soils, Soil Science and Plant Nutrition, 49:1, 61-68, DOI: 10.1080/00380768.2003.10409980

To link to this article: <u>https://doi.org/10.1080/00380768.2003.10409980</u>

d	D	1	1
Е	Г		
Г	Г		
С			

Published online: 22 Nov 2011.



Submit your article to this journal 🗗

Article views: 358



View related articles



Citing articles: 10 View citing articles 🕑

Decomposition and CO₂-C Evolution of Okara, Sewage Sludge, Cow and Poultry Manure Composts in Soils

Michael Miyittah and Kazuyuki Inubushi*

Graduate School of Science and Technology and *Faculty of Horticulture, Chiba University, Matsudo, 271–8510 Japan

Received April 26, 2002; accepted in revised form November 8, 2002

A laboratory experiment was conducted to evaluate the decomposition of various composts in soils by determining the C mineralization rate and microbial biomass level. The differences in the decomposition rate of the composts were examined. The composts used consisted of okara compost made from soymilk residues, cow manure, poultry manure, and sewage sludge composts, which were applied at rates of 50 or 150 g kg⁻¹ soil to a Chiba Light-colour Andosol. The results showed that, in general, the amount of CO_2 -C released increased rapidly at the initial stage, but the pattern differed among the composts used. The production of CO_2 -C depended on the amount applied and the nature of the compost materials. The CO_2 -C evolution from okara-treated soil was much higher than that from the soils treated with the other three composts, presumably due to the higher level of labile C as evidenced by the larger amount of microbial biomass. The value of qCO₂ (CO₂ production per unit microbial biomass) was lower in the okara compost than in the other composts. qCO₂ was linked to the decomposability of organic materials, reflected in the CO₂-C evolution.

Key words: CO_2 -C evolution, decomposition, metabolic quotient (q CO_2), microbial biomass, mineralization.

Separate collection of organic wastes followed by composting and reuse into soil has been developed as the basic principle of wastes disposal in Japan. About 280 million tons of organic residues are generated in a year, accounting for 60% of the total solid wastes produced (Chino 2001). For sustainable agriculture, land application of organic materials is mostly practiced. The decomposition of these materials by microorganisms is a key source of supply of major plants nutrients. Organic materials added to the soils contain a wide range of C compounds that vary in their rates of decomposition. The biological breakdown of the added materials depends on the rate of degradation of each of the C-containing materials present in the sample (Gilmour et al. 1977; Reddy et al. 1980).

The cycling of soil C is essential for nutrients in soil and plant uptake, firstly, because important nutrients such as N and S become mineralized stoichiometrically with C (McGill and Cole 1981) and secondly, because available C is necessary as an energy source for heterotrophic soil microorganisms. Against this background, an incubation experiment was designed to evaluate the effects of various composts on microbial respiration and biomass in soil. The biomass supplied mainly a portion of the total amount of CO_2 mineralized. The rates of CO_2 efflux provide an indication of organic matter quality, and the conduciveness of the soil environment to the decomposition process provides conditions of moisture and temperature that are not limiting (Sparling 1997).

The decomposition of the composts (organic matter) reflects the biological availability of the soil carbon, while the release of CO_2 following the addition of relatively simple substrates indicates the potential carbon-mineralizing capacity of the microflora and has contributed significantly to studies on soil metabolism (Stotzky 1960; Bradley and Fyles 1995).

The okara compost was prepared from soymilk residues and sawdust, which could potentially be useful as soil amendment / organic fertilizer. In this study, the okara compost was compared with sewage sludge compost as well as with cow and poultry manure composts available on the market. The objective was to determine the degree of decomposability of the respective the composts and the amount of carbon mineralized for the increase of the C content of soils as a reflection of their quality, to utilize as a compost the large amount of okara produced each year in Japan.

MATERIALS AND METHODS

Incubation experiment and analytical procedures. An incubation experiment was carried out with four different composts; the okara compost (OK) was obtained from Kikkoman Co-operation, Noda, Chiba, Japan. Sewage sludge (SS) was obtained from the Laboratory of Plant Nutrition, The University of Tokyo, while the cow manure (CM) and poultry manure (PM) composts were commercial products. The composts were dried at 65°C to determine the dry matter content. Fresh compost samples were digested with H₂SO₄ using Kjeldhal techniques for the determination of the total-N contents. Ammonium and nitrate in soil were extracted with 2 M KCl at 1:10 ratio and their contents were determined (Rowell 1994). Total C content in the soil, compost and compost-soil mixture was also determined (Rowell 1994). The pH was determined in a 1:10 compost: water slurry mixture (Rowell 1994). The composition of the Na, K, Mg, Ca, Al, Ni, Cu, and Zn elements in the composts was determined by ICP (ICPS-1000IV, Shimadzu, Kyoto, Japan) (Topper and Kotuby-Amacher 1990).

The soil in which the composts were incubated was a Chiba Light-colour Andosol, classified as Typic Hapludand (US Soil Taxonomy: Soil Survey Staff 1998). The soil was taken from an upland field (0-20 cm) before the application of fertilizer (Table 1). The soils were air-dried and sieved through a 2 mm mesh-sieve.

The incubation was carried out for 40 g (d.w.) of the soil samples mixed with the composts in 200 mL containers and wetted to a moisture content at 60% WHC, to ensure that microbial activities would function at their optimal rate. The amounts of composts added were 0, 2, or 6 g, corresponding to 0, 50, or 150 g kg⁻¹ soil fresh weight. The containers were covered tightly with a thin plastic film (Parafilm), and incubated at 30°C in the dark for 16 weeks. Every week, the moisture content was readjusted by weight to maintain a constant moisture content during the incubation period. Three replicate samples of each treatment were placed in 1-L airtight jars that contained water to maintain a moist atmosphere and a vial with 20 mL 0.3 M NaOH to absorb CO₂. Excess NaOH was titrated with 0.1 M HCl after precipitation of carbonates with BaCl₂, and subtracted from the control without soil to determine the amount of CO_2 evolution from the treated soils during that week. The weekly rates of CO_2 evolution in the compost-treated soils were used to derive the decomposition rates of the composts. The amount of C evolved as carbon dioxide from the composting samples was calculated by subtracting the amount produced by the control soil from that produced by the compost-treated soil. Data concerning CO_2 -C evolution of the compost samples were fitted to kinetic exponential / linear equations.

The amount of microbial biomass carbon was determined by the chloroform fumigation extraction method (Vance et al. 1987). The amount of total extractable C was measured by automated combustion (TOC 5000, Shimadzu). The biomass C level was calculated using the equation: biomass C = 2.64 E_c , where E_c = differences in the contents of organic C of the fumigated and unfumigated samples.

Statistical design and analyses. A simple linear regression analysis was used to correlate the differences between the composts and their rate of decomposition. One-way analysis of variance (ANOVA), as statistical analysis, was carried out. All the analyses were performed using Sigma Stat for Windows 2.03 computing package (Jandel Corporation, Chicago, USA).

RESULTS AND DISCUSSION

Quality of organic manure composts

Organic by-products applied as composts contain a significant amount of proteinaceous and carbonaceous substrates, which can enrich the soil with C, N, and other nutrients. Some chemical properties of the various composts are presented in Tables 2 and 3, respectively. The pH (H₂O) in these composts ranged between 5.4 and 8.9 and EC from 2.2 to 6.1 dS m⁻¹. For the improvement of agricultural soils, compost with pH < 7.2 is required (Rynk et al. 1992), whereas for EC, the acceptable level for a compost is >4.0 dS m⁻¹. The pH (5.4) and EC (4.2) values in OK compared with the values in the other composts satisfied these criteria, except for SS, which had low EC values.

The C / N ratios of the composts were in the increasing order of OK < SS < PM < CM, respectively (Table

	Table 1.	Some proper	ties of the sc	pil.		
Soil	pH (H ₂ O)	$\frac{\text{Mineral N}}{\text{NH}_4^+-\text{N}}$	$\frac{N (g kg^{-1})}{NO_3^{-}-N}$	T-C (g kg ⁻¹)	T-N (g kg ⁻¹)	C / N ratio
Chiba Light-colour Andosol*	4.87	30.7	75.4	64.9	6.86	9.46

*Typic Hapludand (US Soil Taxonomy: Soil Survey Staff 1998).

2). The C / N ratio has been used to indicate the stability of the composts. Ratios below 15 were assumed to be indicative of a stable and mature compost. However, Mathur (1991) reported that a mature compost should have a C/N ratio of about 10 as in humus, whereas, composts with a high C/N ratio are often associated with plant phytoxicity and with the presence of volatile fatty acids. The values of C / N reported ranged from 5.8 to 8.3 (Table 2). These values satisfied the criteria outlined above. Carbon/nitrogen ratios are commonly used to predict the mineralization potential of organic materials (Azmal et al. 1996). In general, the higher the C / N ratio, the slower the mineralization. It was therefore expected in this study that the mineralization rate would be higher in the soils treated with composts with a low C / N ratio and in the soils treated with OK (C / N ratio of 5.8) than in those treated with CM (C/N ratio of 8.3).

Carbon mineralization

Decomposition of the four composts in the soil, as determined by CO_2 evolution, was directly related to the amount of carbon in the compost-soil mixtures (Fig. 1).

The amount of cumulative CO_2 -C mineralized appeared to follow a first order kinetics for all the treatments. All of the compost-mixtures released CO_2 in relation to the total amount present, starting with a release of C during the first week of incubation, with decreasing rates thereafter. In estimating the decomposition rates of the various organic C pools in each organic material, the rates were taken from the slopes of the curves obtained from their natural logarithms.

A steep slope represents the highest decomposing rate, which was considered to be the most readily decomposable organic C fraction. The application of compost at 50 and 150 g kg⁻¹ rates gave values of 3.07 and 3.43 for OK, 2.93 and 2.96 for PM, 1.72 and 2.05 for CM, and 1.44 and 1.78 μ g C g⁻¹ week⁻¹ for SS, respectively. Compost application at a rate of 100 g kg⁻¹ resulted in a higher level of CO₂ evolution, as indicated in all the compost-treated soils. The results showed a higher level of CO₂ evolution for OK than for the other composts. The amount of CO₂ decreased in the order of OK > PM > SS > CM. The higher level of OK may therefore suggest that higher rates of decomposition took place, which reflected the presence of available C

 Table 2.
 Chemical properties of the composts used.

Commente	pH (H ₂ O)	pH (KCl)	Cl) EC	Mineral N	(mg kg ⁻¹)	T-C	$\frac{\text{T-N}}{(\text{g kg}^{-1})}$	C / N ratio
Composts	1:10	1:10	(dS m ⁻¹)	NH4 ⁺ -N	NO ₃ N	$(g kg^{-1})$		
Okara (OK)	5.4	5.3	4.2	219	9.54	514	88.2	5.83
Sewage sludge (SS)	6.5	ND	2.2	747	28.14	288	44.1	6.53
Cow manure (CM)	6.5	6.3	3.7	109	54.84	157	18.9	8.31
Poultry manure (PM)	8.91	8.30	6.1	50.6	15.18	251	35.1	7.15

ND: Not determined.



Fig. 1. Rates of CO₂ evolution from okara, cow manure, poultry manure, and sewage sludge composts incubated in soil Mean and standard error of three replicates is shown. -*- control, - \Box - okara compost@5%, -**e**- okara compost@15%, - \diamond - cow manure@5%, -**e**- cow manure@15%, - \diamond - poultry manure@15%, - \diamond - sewage sludge@5%, -**e**- sewage sludge@15%.

from readily decomposable compounds such as protein, as reported by Lerch et al. (1992). The lower decomposition value of CM may indicate the predominance of less decomposable substances that would, therefore, be the most resistant to microbial decomposition.

As shown in Table 3 for the Ni, Cu, and Zn contents, even though the Cd content was not reported in this study, Sadovnikova et al. (1995) indicated that the presence of heavy metals such as Cd, Ni, Cu, Pb, and Zn exerts an inhibitory effect on decomposition. For the total Ni or Cu and Zn concentrations, the acceptable concentrations for composts were <400, <1,500, and <1,800 mg kg⁻¹, respectively (Rao and Pandey 1996). The Cu and Zn concentrations ranged from 0.8 to 6.5 and 4.1 and 19.7 mg kg⁻¹, respectively whereas, that of Ni from 0.2 to 0.4 mg kg⁻¹ was almost negligible. These levels were far too low among the composts used (Table 3), indicating the overall suitability and quality of the composts as soil amendment.

The results of the linear regression analyses to select the optimum prediction of the amount of C mineralized during the study are presented in Fig. 2. A linear equation was drawn to represent delta (Δ) CO₂ evolution, which is equivalent to the difference between the CO₂ evolution from soil treated with 50 and 150 g kg⁻¹ compost, as indicated by the following equations. For OK, $\Delta \text{CO}_2 = 1.54t - 1.88$; PM: 0.67t + 1.34; SS: 0.388t + 3.17; and CM: 0.312t + 3.11, with a statistical difference among the composts at p < 0.001 (Fig. 2), where t in the equation represents the time of incubation (week). The order of decomposition was OK > PM > SS > CM. To obtain more details in the plots for OK, the period 2–8 weeks was examined and it showed some irregularities; the plots were lower than the regression line, which is discussed in the next section.

The Δ CO₂-C evolution represented a zero-order model if it is assumed that all the materials were undergoing decomposition at the same level (Fig. 2). Actually, this assumption may not reflect all the decomposable fractions, since the composts were made of different feedstocks and the compounds may be resistant to microbial attack, hence the use of other stepwise models might be preferable. However, the equation still supports the prediction of low rates of decomposition among the composts (Lerch et al. 1992).

These results may further be used to justify the approach of the simulation models for the C–N turnover of organic residues, where the rates of decomposition of organic C and N in soil are based on the properties of decomposing materials, C / N ratio, and other soil properties, as suggested by Van Veen et al. (1985), Molina et al. (1990), and Hadas and Portnoy (1994).

Table 3. Elemental composition of okara, sewage sludge, cow manure, and poultry manure composts.

Composts / elements (mg kg ⁻¹)	Mg	К	Ca	Р	Na	Al	Ni	Cu	Zn
Okara	31.4	216	77	49	13.2	5.4	ND	0.8	4.1
Sewage sludge	52.3	129	115	158	15.4	156.2	0.4	6.5	14.9
Cow manure	50.0	128	390	76	23.6	156.3	0.4	4.0	19.7
Poultry manure	55.0	130	395	202	125.3	20.9	0.2	2.8	18.6

ND: Not determined.



Fig. 2. Δ CO₂ evolution with time. Mean and standard error of three replicates is shown. \blacksquare okara (OK), \blacktriangle sewage sludge (SS), \square cow manure (CM), \bullet poltry manure (PM).

Microbial biomass

Microbial biomass has been used as an indicator of soil quality, as it responds rapidly to the changes in the soil environment that can be associated with soil contamination. The nutrients in microbial biomass are potentially available to plants (Smith and Paul 1991; Chander et al. 1995). Under favorable conditions, microbial biomass is positively correlated with the contents of organic matter, and N and P in the biomass, which are also positively correlated with N, P, and C availability (He et al. 1997).

The microbial biomass C showed similar trends for

each of the composts CM, SS, PM, and OK (Fig. 3). There was a gradual increase from two weeks to about 6 weeks, followed by a general decreasing trend for PM, SS, and CM. On the other hand, OK showed a sharp rise from 2 weeks to about 8th weeks, followed by a sharp decline to the 12th week, and then a gradual decrease to the 16th week. At the end of incubation, OK contained a higher amount of accumulated biomass C than the other composts, which all showed the accumulation at almost the same level, while the accumulation was slightly higher than that of the control. Some irregularities have been observed in ΔCO_2 between 2–8 weeks (Fig. 2),



Fig. 3. Microbial biomass C of compost at 150 g kg⁻¹ application rate: Bars denote mean and SE of three replicates. -o- control, -**a**- okara, -**a**- sewage sludge, -**a**- cow manure, -**b**- poultry manure.



Fig. 4. Relationship between metabolic quotients (qCO_2) and incubation time: $qCO_2(OK)$, $qCO_2(CM)$, $qCO_2(PM)$, $qCO_2(SS)$ denote metabolic quotients for okara, cow manure, poultry manure, and sewage sludge composts, respectively. The lines represent the regression equations for the composts. $\blacksquare qCO_2(OK)$, $\Box qCO_2(CM)$, $\blacklozenge qCO_2(PM)$, $\blacktriangle qCO_2(SS)$.

which may account for the amount of biomass built up during this period.

The differences in the compost fractions probably reflected the differences in the growth and activity of the succession of microbial species, which are largely influenced by the concentration of readily available C in the substrates. As reported by Sakamoto and Hodono (2000), microbial biomass C shows a strong positive correlation with the turnover time of biomass C, since the turnover time is closely related to the survival of microorganisms. Kouno et al. (2002) also observed the increase in the total biomass C pool size and turnover time as a result of the addition of ryegrass to soil. The higher level of biomass C for OK shown in Fig. 3 at a 150 g kg⁻¹ application rate than in CM could be due to the activity of various microbial species, availability of easily decomposable substrates and their efficiency in decomposition. The observation of qualitative differences between microbial communities can be associated with differences in the microbial species. As for the selective adaptation of microbial populations to certain plant residues / composts, no conclusive evidence can be found in the literature (Verstraete and Voets 1977). The soil microbial biomass is thus involved in the decomposition of organic materials and cycling of nutrients in soil. Therefore, it appears that the mineralization potential of OK is slightly higher than that of the other composts, as evidenced by the lower C / N ratio (Table 2).

Metabolic quotients (qCO_2)

The soil microbial biomass, as living microbial cells in soil, is the main agent responsible for CO_2 evolution. The ratio of CO_2 evolution to microbial biomass is designated as metabolic quotient: qCO_2 (Anderson and Domsch 1986). The qCO_2 , or specific respiration rate, has been used in soil microbial biomass analyses for investigations on the maintenance of energy (Anderson and Domsch 1985a, b). The metabolic quotient is high when the microorganisms cannot fully metabolize C efficiently while feeding on N. Similarly, a higher qCO_2 also reflects the microbial response to adverse environmental conditions or disturbances (Wardle and Ghani 1995).

The metabolic efficiency of microbial biomass is an indication of the availability of organic C and its utilization. The qCO₂, therefore, evaluates the efficiency of soil microbial populations in utilizing organic C compounds. In this study, the qCO₂ decreased exponentially in the order of CM > SS > PM > OK, with (p < 0.05) and R^2 values of 0.95, 0.77, 0.75, 0.20, respectively (Fig. 4). A lower value was observed in the case of OK. Anderson and Domsch (1993) showed that a lower prevailing soil pH had a negative effect on qCO₂. This might explain the observation in OK, although in our

data, a moderately high soil pH in CM, SS, and PM gave seemingly corresponding qCO_2 values. Comparatively, the metabolic quotient was higher in CM than in OK, implying that the microbes were under stress while utilizing CM and, therefore, could not fully metabolize C efficiently. This behaviour in microbial activities could be linked to the mineralization pattern of organic materials such as compost. The order of decomposition as indicated by the metabolic quotients was similar to that observed by the use of the C / N ratio in predicting rates. The observed qCO₂ was linked to the decomposability of the organic materials, as indicated in the CO₂-C evolution.

Therefore, the use of the metabolic quotient may also enable to predict the decomposability of organic materials such as compost, with the additional advantage of knowing the microbial actions involved in terms of biomass. The biomass is the most important indicator of microbial performance in soil, especially in combination with CO_2 production (Anderson and Domsch 1993). The diversity and composition of microbial populations should be taken into account to further strengthen q CO_2 actions, as indicated by the fact (Mamilov and Dilly 2002) that bacterial activities during decomposition built up microbial biomass. Soils with a heavy metal history should also be taken into consideration when calculating q CO_2 values, since heavy metal actions affect the metabolic activities of microorganisms (Insam et al. 1996).

While qCO_2 undoubtedly reflected the microbial efficiency for carbon utilization in this short-term study, Zaman et al. (2002) reported its inconsistency in long-term evaluation of organic wastes due to conflicting results in some studies. Although Zaman et al. (2002) did not discuss the cause of this phenomenon, this could be due to the fact that in a long-term evaluation, the microbial nutrient status changes considerably, depending on other environmental factors, the composition and the physiological state of the microflora, and the availability of decomposable substrates. However, qCO_2 could possibly be used as microbial indicator in the short term.

The soil microbial activity most probably depends on the biomass level and metabolic quotient. However, there was an inverse relationship between the biomass level in soil and the metabolic quotient (Santruckova and Straskraba 1991). This, therefore, suggests that, in some cases, higher levels of carbon inputs to the soil increase the microbial biomass but not the metabolic activity, which in turn leads to a decrease of the metabolic quotient (Insam et al. 1991).

Our results indicate that parameters such as microbial biomass and metabolic quotient are indices that affect the decomposability and carbon utilization of organic materials such as composts. The composts therefore provided additional C for microbial respiration and growth, but the fraction of decomposed C used for microbial efficiency might have been larger with a larger C supply, and qCO_2 plays a significant role in C efficiency, utilization, mineralization and in macroaggregate formation, as discussed by Miller and Dick (1995).

The addition of organic materials with easily degradable C compounds to soil appears to be suitable for stabilizing and supplying carbon and very useful from the viewpoint of C sequestration and ways of maintaining or increasing the humus content of soils. The order of qCO_2 seems to follow that of C and N mineralization.

Acknowledgments. This work was partly supported by Kikkoman Co-operation for the supply of okara compost and also the assistance extended by M. Matsuura and Y. Hanya of the Kikkoman Co-operation Research Division Unit is fully acknowledged.

REFERENCES

- Anderson TH and Domsch HK 1985a: Maintenance of carbon requirements of activity-metabolizing microbial populations under in situ conditions. *Soil Biol. Biochem.*, **17**, 197–203
- Anderson TH and Domsch HK 1985b: Determination of ecophysiological maintenance carbon requirements of soil microorganisms in a dormant state. *Biol. Fertil. Soils*, 1, 81-89
- Anderson TH and Domsch HK 1986: Carbon assimilation and microbial activity in soil. Z. Pflanzenernaehr. Bodenkd., 149, 457-468
- Anderson TH and Domsch HK 1993: The metabolic quotient for CO_2 (q CO_2) as a specific activity parameter to assess the effects of environmental conditions, such as pH, on the microbial biomass of forest soils. *Soil Biol. Biochem.*, **25**, 393–395
- Azmal AKM, Marumoto T, Shindo H, and Nishiyama M 1996: Mineralization and microbial biomass formation in upland soil amended with some tropical plant residues at different temperatures. Soil Sci. Plant Nutr., 42, 436–473
- Bradley RL and Fyles JW 1995: A kinetic parameter describing soil available carbon and its relationship to rate increase in C mineralization. *Soil Biol. Biochem.*, **27**, 167–172
- Chander K, Brookes PC, and Harding SA 1995: Microbial biomass dynamics following addition of metal-enriched sewage sludges to a sandy loam. *Soil Biol. Biochem.*, **27**, 1409– 1421
- Chino M 2001: Status of organic recycling/composting in Japan. *BioCycle*, **42**, 61–62
- Gilmour CM, Broadbent FE, and Beck SM 1977: Recycling of carbon and nitrogen through land disposal of various wastes.
 In Soils for Management of Organic Wastes and Waste Waters, Ed. LE Elliott and FJ Stevenson, p. 173-294, Soil Sci. Soc. Am., Madison, WI
- Hadas A and Portnoy R 1994: Nitrogen and carbon mineralization rates of composted manures incubated in soil. J. Environ. Qual., 23, 1184–1189

- He Z, Yao L, Chen G, Zhu J, and Huang C 1997: The relationship of crop yield to microbial biomass in highly weathered soils of China. *In* Plant Nutrition for Sustainable Food Production and Environment, Ed. T Ando et al., p. 745–746, Kluwer Academic Publishers, Tokyo
- Insam H, Hutchinson TC, and Reber HH 1996: Effects of heavy metal stress on metabolic quotient of the soil microflora. *Soil Biol. Biochem.*, 28, 491–694
- Insam H, Mitchel CC, and Dormaar, JF 1991: Relationship of soil microbial biomass and activity with fertilization practice and crop yield on three Ultisols. *Soil Biol. Biochem.*, 23, 459–464
- Kouno K, Wu J, and Brookes PC 2002: Turnover of biomass C and P in soil following incorporation of glucose or ryegrass. *Soil Biol. Biochem.*, 34, 617–622
- Lerch RN, Barbarick KA, Sommers LE, and Westfall DG 1992: Sewage sludge proteins as labile carbon and nitrogen sources. Soil Sci. Soc. Am. J., 56, 1470–1476
- Mamilov AS and Dilly OM 2002: Soil microbial eco-physiology as affected by short-term variations in environmental conditions. Soil Biol. Biochem., 34, 1283–1290
- Mathur SP 1991: Composting process. In Bioconversion of Waste Materials to Industrial Products, Ed. AM Martin, p. 147-186, Elsevier, New York
- McGill WB and Cole CV 1981: Comparative aspects of cycling of organic carbon, nitrogen, sulfur, and phosphorous through soil organic matter. *Geoderma*, **26**, 267–286
- Miller M and Dick RC 1995: Dynamics of soil C and microbial biomass in whole soil and aggregates in two cropping systems. Appl. Soil Ecol., 2, 253–261
- Molina JAE, Hadas A, and Clapp CE 1990: Computer simulation of nitrogen turnover in soil and priming effect. Soil Biol. Biochem., 22, 349–353
- Rao BSM and Pandey SP 1996: Aerobic thermophilic composting of piggery solid wastes. *Water Sci. Technol.*, **33**, 89–94
- Reddy KR, Khaleel R, and Overcash MR 1980: Carbon transformations in land areas receiving organic wastes in relation to nonpoint source pollution: A conceptual model. J. Environ. Qual., 9, 434–442
- Rowell DL 1994: The meaning of pH and its measurement, the determination of organic nitrogen and the dichromate method for the determination of oxidizable carbon and soil organic matter. *In* Soil Science, Methods and Applications, p. 48–161, Addison Wesley Longman Limited, London, UK
- Rynk R, van de Kamp M, Willson GB, Singley ME, Richard TL, Kolega JL, Gouin FR, Lalibertry L Jr, Day K, Murphy DW, Hoitink HAJ, and Brinton WF 1992: On-Farm Composting Hand-Book, p. 186–187, NRAES, Cornell University, Ithaca, New York
- Sadovnikova LK, Otabbong E, Iakimenko OS, Nilsson L, and Persson J 1995: Dynamic transformations of sewage sludge and farmyard manure components: 2. Copper, lead and cadmium forms in incubated soils. Agric. Ecosyst. Environ., 58, 127-132
- Sakamoto K and Hodono N 2000: Turnover time of microbial biomass carbon in Japanese upland soils with different textures. *Soil Sci. Plant Nutr.*, **46**, 483–490
- Santruckova H and Straskraba M 1991: On the relationship between specific respiration activity and microbial biomass in soils. *Soil Biol. Biochem.*, **23**, 525–532

- Smith JL and Paul EA 1991: The significance of soil microbial biomass estimations. In Soil Biochemistry, Vol. 6, Ed. JM Bollag and G Stotzky, p. 359–369, Marcel Dekker, New York
- Soil Survey Staff 1998: Keys to Soil Taxonomy 8th Ed., p. 326, USDA National Resources Conservation Service, Madison, WI
- Sparling GP 1997: Soil microbial biomass, activity and nutrient cycling as indicators of soil health. *In* Biological Indicators of Soil Health, Ed. C Pankhurst, BM Doube, and VVSR Gupta, p. 97–119, CAB International, Wallingford
- Stotzky G 1960: A simple method for the determination of respiratory quotient of soils. *Can. J. Microbiol.*, **6**, 439-452
- Topper K and Kotuby-Amacher J 1990: Evaluation of closed vessel acid digestion method for plant analyses using inductively coupled plasma spectrometry. *Commun. Soil Sci. Plant Anal.*, **21**, 1437–1455

Van Veen JA, Ladd JN, and Amato A 1985: Turnover of carbon

and nitrogen through the microbial biomass in a sandy loam and clay soil incubated with ¹⁴[C(U)] glucose and [¹⁵N]-(NH₄)₂SO₄ under different moisture regimes. *Soil Biol. Biochem.*, **17**, 747–756

- Vance ED, Brookes PC, and Jenkinson DS 1987: An extraction method for measuring microbial biomass C. Soil Biol. Biochem., 19, 703–707
- Verstraete W and Voets JP 1977: Soil microbial and biochemical characteristics in relation to soil management and fertility. *Soil Biol. Biochem.*, 9, 253–258
- Wardle DA and Ghani A 1995: A critique of the microbial metabolic quotient (qCO_2) as a bioindicator of disturbance and ecosystem development. *Soil Biol. Biochem.*, **27**, 1601– 1610
- Zaman M, Di J, Sakamoto K, Goto S, Hayashi H, and Inubushi K 2002: Effects of sewage sludge compost and chemical fertilizer application on microbial biomass and N mineralization rates. Soil Sci. Plant Nutr., 48, 195-201